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## **Vitamin D receptor polymorphisms relate to risk of adenomatous polyps in a sex specific manner.**

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**Short Title:** VDR genotypes and risk of adenomatous polyps

**Key Words:** vitamin D, adenomatous polyps, *VDR*, polymorphisms, nutrigenetics.

## Abstract

**Background/Aims:** Vitamin D receptor (VDR) gene polymorphisms may influence risk for adenomatous polyps, a benign precursor to colon cancer, via modulation of vitamin D sensitive pathways, including cell proliferation and differentiation. However, results have been mixed and any association remains contentious. Failure to clinically exclude the presence of adenomatous polyps in control cohorts may contribute to the lack of consensus. Therefore, we assessed the role of the FokI, BsmI, ApaI and TaqI *VDR* polymorphisms in modifying risk for adenomatous polyps, adjusting for a range of dietary and lifestyle variables. **Methods:** Blood was collected from colonoscopy patients (n=258) and *VDR* polymorphisms assessed by RFLP. Dietary habits were estimated from food frequency questionnaires. Odds ratios for adenomatous polyps were calculated by genotype, stratified by sex, and adjusted for age, lifestyle and dietary factors. **Results:** FokI was associated with modified risk for AP in males, whilst the BsmI/ApaI/TaqI haplotype was associated with modified risk in females. No interaction was found between *VDR* variants and vitamin D intake. **Conclusion:** This study offers novel insight into the potential for *VDR* genetics to contribute to risk for adenomatous polyps, and is the first to demonstrate a sex-specific relationship between these polymorphisms and risk for adenomatous polyps.

## Introduction

Adenomatous polyps (AP) are the benign precursor to colorectal cancer (CRC), a cancer with multiple known modifiable dietary risk factors (1-3). Vitamin D may contribute to the aetiology of AP (4-6) via modulation of multiple vitamin D sensitive pathways that can influence tumourigenesis, including angiogenesis, cell proliferation and differentiation, immunomodulation, and protection from oxidative stress (7-9). This may occur indirectly via regulation of calcium homeostasis, or via direct interaction with the vitamin D receptor (VDR) (7). *VDR* polymorphisms may influence these pathways and modify risk of disease.

Numerous polymorphisms have been identified in the *VDR* gene. Of these only the FokI (rs2228570, previously rs10735810) polymorphism is known to generate a modified protein (10). The “F” allele has a later start codon than the “f” allele, generating a shorter more transcriptionally active protein (10-12). Three well studied polymorphisms are located at the 3’ end of the gene; BsmI (rs1544410) and ApaI (rs7975232) (13-15), and TaqI (rs731236) (16). These polymorphisms are in high linkage disequilibrium forming a haplotype block (13, 16-18). These variants do not result in changes to the final VDR protein, but they have been linked to altered bone density (19) and risk of multiple diseases (20-22). This may occur via altered mRNA stability (11, 16) or these may be silent marker polymorphisms for other highly linked functional polymorphisms (23), such as a variable length of the poly (A) tail. The “baT” haplotype, containing the ancestral version of each allele, is linked to the long poly (A) variant, while the polymorphic “BAt” haplotype is linked to the short variant (16, 24).

Several studies have investigated the role of common *VDR* polymorphisms in the risk of CRC, however results have varied and the association remains contentious. Some studies have found no association between FokI status and CRC (25, 26), while others have found varying associations. Carriage of the “f” allele reportedly increased risk in a Chinese-Singaporean population, (27) and in those with high energy intake, low physical activity or obesity (28). Conversely, others have reported that carriage of the “F” allele increases CRC risk (4, 29, 30). This has been demonstrated in those with the “baT” haplotype (29), and regardless of BsmI genotype and poly (A) tail length, (30) suggesting that the FokI

haplotype may exert a stronger influence than some other polymorphisms. Consideration of the BsmI polymorphism alone has also yielded mixed results; both an increased (25, 31) and reduced risk (32, 33) in the presence the “B” allele, and no association with CRC (28, 34-36) have been reported. A 2014 meta-analysis found reduced risk in the presence of the “b” allele (37).

These discrepancies may be explained by failure to clinically exclude the presence of AP in control subjects. Controls for CRC studies are often recruited from licence and insurance registers and colonoscopies are not commonly performed (25, 27, 28, 30, 34). Only limited studies have examined the influence of *VDR* polymorphisms on AP risk in colonoscopy verified cases and controls; therefore it is difficult to draw firm conclusions. Some have found no relationship for multiple *VDR* polymorphisms (38, 39), BsmI (40), TaqI (41) or FokI (42). While Ingles *et al.* found that BsmI and FokI did not influence risk of AP alone, presence of the “f” allele was associated with increased risk of large (>1cm) polyps (18). A small 2011 meta-analysis found no association between FokI or BsmI genotype and AP risk (6). These discrepancies may be explained by differences in other dietary and lifestyle risk factors included in the analyses, or interactions between genotypes and nutritional status in the absence of direct association with genotype alone (43, 44).

Combined analysis of the BsmI/ApaI/TaqI haplotype and FokI are yet to be conducted in a single cohort using colonoscopy confirmed AP as cases and confirmed absence of polyps as controls. Therefore, we assessed the role of these four common *VDR* polymorphisms in modifying the risk for AP in a colonoscopy confirmed cohort, applying adjustment for a range of dietary and lifestyle factors.

## **Methods**

### ***Subjects and sample collection***

Patients (n=263) undergoing routine screening for colonic pathology at a gastroenterology clinic (Gosford Hospital, NSW, Australia), aged 18-89 were recruited. 258 participants (43.80% male) who completed the required food frequency questionnaire, gave blood samples for DNA isolation, and received a definitive diagnosis as to the presence or absence of AP were included in this analysis.

Informed consent was obtained prior to participation under University of Newcastle Human Research Ethics Committee approval number H-429-0407.

### ***Food frequency questionnaires***

Daily intake of macro and micronutrients were estimated using a validated interviewer administered food frequency questionnaire, covering 225 food items and all food groups. Subjects also provided a list of all supplements and their frequency of intake. The food frequency questionnaires were analysed using Foodworks™ (Version 2.10.146, Xyris Software, Brisbane, QLD, Australia) (45, 46).

### ***Genotyping***

Genomic DNA was isolated from whole blood and amplified using PCR. Restriction fragment length polymorphism (RFLP) assays and gel electrophoresis were used to genotype four common polymorphisms of the *VDR* gene; BsmI, ApaI, TaqI and FokI (45). The presence of restriction sites were coded with lower case letters (BsmI- 'b'; ApaI- 'a'; TaqI- 't'; FokI- 'f'), and the absence of restriction sites as the same letters in upper-case (BsmI- 'B'; ApaI- 'A'; TaqI- 'T'; FokI- 'F'). See supplementary methods for additional details. Haplotypes were reconstructed using PHASE v2.1(47, 48).

### ***Statistics***

Statistical analyses were performed using JMP (Version 11, SAS Institute Inc., Cary, NC, USA). Relationships were analysed by standard least squares or nominal logistic regression analysis, or Chi-squared tests, as appropriate. Analyses were stratified by sex and adjusted at a minimum for age, smoking history and reported alcohol intake. Additional models were performed adjusting for intake of energy, and a range of micro- and macro-nutrients previously linked to AP or CRC risk (total energy, dietary fibre, fat, folate, vitamin D, calcium, and iron intakes) and adjusted odds ratios (OR) with 95% confidence intervals (CI) were calculated. Outcomes were considered to be statistically significant at  $p \leq 0.05$ . Models were conducted with crossed terms to investigate interactions between variables where

appropriate. Descriptive statistics (mean±SEM, range) were calculated by sex and presented as required. Due to the restricted sample size, genotypes were combined to allow analysis by presence vs. absence of the ancestral or polymorphic alleles, as appropriate. Hardy-Weinberg equilibrium was tested for each polymorphism using a  $\chi^2$  test.

## Results

There were several significant differences between cases and controls, and male and female cohorts (Table 1). In females, cases were significantly older than female controls (65.50±1.71 vs. 59.25±1.08 years, p=0.003). In males, cases consumed significantly less calcium (844.64±63.39 vs. 1112.21±73.36 mg/day, p=0.007) than controls. Somewhat surprisingly, male controls were more likely to have a history of smoking than cases ( $\chi^2= 4.4$ , p=0.04; Table 1). Males (both cases and controls) consumed significantly more alcohol (2.00±0.50 vs. 0.69±0.25 g/day, p=0.02; 1.89±0.25 vs. 0.57±0.10 g/day, p=0.001, respectively), and more kilojoules (10167±459 vs. 8693±568 kJ/day, p=0.05; 10670.60±441.30 vs. 9013.28±340.61 kJ/day, p=0.003, respectively) than female cases. Male cases consumed significantly less calcium (844.64±63.39 vs. 1304.88±213.80 g/day, p=0.05), and significantly more iron (15.97±0.97 vs. 12.76±8.91 g/day, p=0.01), than female cases. Male controls were significantly older (63.82±1.45 vs. 59.25±1.08 years, p=0.01), were more likely to be smokers ( $\chi^2= 4.6$ , p=0.03), and consumed significantly more fat (96.49±5.38 vs. 80.84±3.11 g/day, p=0.01) relative to female controls (Table 1).

The allele and genotype frequencies for the tested polymorphisms are given in Table 2. The genotype frequencies did not deviate from Hardy-Weinberg equilibrium expectations (FokI p=0.28; BsmI p=0.63; ApaI p=0.91; TaqI p=0.56). As expected, BsmI, ApaI and TaqI demonstrated high linkage disequilibrium in this cohort (Supplementary Figure 1), all with D' statistics over 0.95 (Supplementary Table 1). These results are comparable to those obtained in the HapMap (<http://hapmap.ncbi.nlm.nih.gov/>, (49)) and 1000 Genomes (<http://www.1000genomes.org/data>, (50)) CEU populations (Centre d'Etude du Polymorphisme Humain (CEPH) Utah residents with ancestry

from northern and western Europe), which have  $D'$  values of 0.98-1.0. There was no evidence of linkage disequilibrium between FokI and any of the other polymorphic sites (Supplementary table 1).

Different alleles were related to an altered risk of AP in males and females (Table 3). In females, presence of the ancestral alleles for BsmI (“b”) and TaqI (“T”) significantly reduced the risk for AP (BsmI unadjusted OR= 0.23, 95% CI= 0.09-0.63,  $p=0.005$ ; TaqI unadjusted OR= 0.25, 95% CI= 0.09-0.69,  $p=0.008$ ). In males, presence of the polymorphic allele for FokI (“F”) significantly increased risk for AP (unadjusted OR= 5.65, 95% CI= 1.07-104.95,  $p=0.043$ ). Adjustment for age, smoking history and alcohol consumption (Model 1; Table 3) and further adjustment for vitamin D and calcium intake (Model 2; Table 3), energy, dietary fibre, fat, folate, and iron (Model 3; Table 3) did not alter the significance of these relationships. No associations were found between AP risk and ApaI alleles in either sex (Table 3).

As BsmI/ApaI/TaqI form a haplotype block (13, 16-18), analysis was repeated with reconstructed haplotypes. Haplotype frequencies are shown in Table 4. The most common haplotype, “baT” (45.16%), contains the ancestral version of each allele (restriction sites for BsmI/ApaI, no restriction site for TaqI). The second most common haplotype “BAt” (30.62%), contains the polymorphic version of each allele (no restriction sites for BsmI/ApaI, restriction site for TaqI). Analysis was repeated for risk of AP by presence of these two common haplotypes. Consistent with the single allele analysis, presence of the “baT” haplotype reduced risk for AP in females (unadjusted OR=0.27, 95% CI=0.10-0.74,  $p=0.01$ ; Table 5). Results remained significant following adjustments (Table 5). No association was found in males.

Combining analyses for both the BsmI/ApaI/TaqI haplotype and the FokI polymorphism revealed that risk was decreased in females with the baTf combination after adjustment for age, smoking history and alcohol consumption (Model 1 OR=0.35, 95% CI=0.11-0.99,  $p=0.05$ ; Table 6), but increased in females with the BA Tf combination (Model 1 OR=2.78, 95% CI=1.08-8.19,  $p=0.05$ ; Table 6), suggesting that the haplotype block is a bigger factor in deciding risk than the FokI allele. Again, results remained significant following adjustment, with the exception of model 3 in the presence of the BA Tf

combination. No statistically significant results were found in males (Table 6). No significant interaction was found between reported vitamin D or calcium intake and any allele or haplotype studied ( $p>0.05$ ).

## **Discussion**

In this Australian cohort the *VDR* polymorphism significantly influenced the risk of AP in a sex specific manner. The start codon FokI polymorphism significantly altered risk in males, whilst the BsmI/ApaI/TaqI haplotype block significantly altered risk in females. Factors contributing to the sex specific associations may include differences in the hormonal milieu and in overall incidence and age of incidence between the sexes. Information on BMI and energy excess are not available in this population and these are likely to differ between males and females. These factors may interact with genetic and hormonal differences to alter risk.

The data presented here supports previous data from Boyapti *et al.* (43) who, in a similar sized study, suggested that high blood calcium levels may reduce risk of AP, particularly in those with at least one “b” allele. However, it is in conflict with the data of Kim *et al.* (40) who found no influence of BsmI genotype, except in those with low serum vitamin D and/or calcium intake. This study is limited in that vitamin D status only represents reported dietary and supplementary intake, and not serum vitamin D levels. Calcium and vitamin D homeostasis may be modulated by *VDR* polymorphisms, and this may be a potential mechanism for altered risk (51, 52). Additional factors may influence systemic levels of vitamin D, calcium and other micronutrients including sun exposure, skin pigmentation, variance in vitamin D binding protein, and other genetic factors, that influence vitamin D and calcium homeostasis. It is also possible that dietary vitamin D and systemic vitamin D may have different influences on AP risk. No interaction was found between the polymorphisms studied and vitamin D or calcium intake. This may suggest that the mechanism of altered risk occurs via an indirect mechanism, or a larger cohort may be required to adequately identify an interaction.

The data presented here is in conflict with that of Slattery *et al.* (28) and Wong *et al.* (27), who found that the “f” allele increased risk of CRC, and supports the data of Sweeney (30), Ingles (18) and Park

(29), who found that the “F” allele increased risk of CRC. Differences between dietary and lifestyle habits of cohorts from different demographics and socio-cultural backgrounds may explain these discrepant results. However, potential differences in risk factors for AP incidence as opposed to the progression to CRC need to be considered. For example, whilst high folate levels may protect against initial occurrence of neoplasms via its essential role in DNA synthesis and repair processes, high folate levels after the initial occurrence may encourage tumour growth via enhanced opportunity for proliferation (53, 54). Differential roles for vitamin D (from diet and UV exposure) and other micronutrients may also exist in the aetiology of AP compared to its sequela, CRC. Additional studies are needed to determine if this is the case and to determine the biological processes involved. Despite the limitations identified, this study offers novel insight into the nutrigenetic risk factors for AP, and is the first to demonstrate a sex specific relationship between these *VDR* polymorphisms and risk for AP.

### **Conflict of Interest Declaration**

All authors declare that they have no conflicts of interest.

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### **References**

- 1 Giovanniucci E: Modifiable risk factors for colon cancer. *Gastroenterol Clin North Am* **31**, 925-43, 2002
- 2 Neugut AI, Garbowski GC, Lee WC, Murray T, Nieves JW, et al.: Dietary risk factors for the incidence and recurrence of colorectal adenomatous polyps. A case-control study. *Ann Intern Med* **118**, 91-5, 1993
- 3 Research. WCRFAIfC. *Continuous Update Project Report. Food, Nutrition, Physical Activity, and the Prevention of Colorectal Cancer.*, 2011.
- 4 Murtaugh MA, Sweeney C, Ma KN, Potter JD, Caan BJ, et al.: Vitamin D receptor gene polymorphisms, dietary promotion of insulin resistance, and colon and rectal cancer. *Nutr Cancer* **55**, 35-43, 2006. doi: 10.1207/s15327914nc5501\_5
- 5 Grau MV, Baron JA, Sandler RS, Haile RW, Beach ML, et al.: Vitamin D, calcium supplementation, and colorectal adenomas: results of a randomized trial. *J Natl Cancer Inst* **95**, 1765-71, 2003

- 6 Lee JE: Circulating levels of vitamin D, vitamin D receptor polymorphisms, and colorectal adenoma: a meta-analysis. *Nutr Res Pract* **5**, 464-70, 2011. doi: 10.4162/nrp.2011.5.5.464
- 7 Deeb KK, Trump DL, Johnson CS: Vitamin D signalling pathways in cancer: potential for anticancer therapeutics. *Nat Rev Cancer* **7**, 684-700, 2007. doi: 10.1038/nrc2196
- 8 Chakraborti CK: Vitamin D as a promising anticancer agent. *Indian J Pharmacol* **43**, 113-20, 2011. doi: 10.4103/0253-7613.77335
- 9 Veldhoen M, Ferreira C: Influence of nutrient-derived metabolites on lymphocyte immunity. *Nat Med* **21**, 709-18, 2015. doi: 10.1038/nm.3894
- 10 Kostner K, Denzer N, Muller CS, Klein R, Tilgen W, et al.: The relevance of vitamin D receptor (VDR) gene polymorphisms for cancer: a review of the literature. *Anticancer Res* **29**, 3511-36, 2009
- 11 Whitfield GK, Remus LS, Jurutka PW, Zitzer H, Oza AK, et al.: Functionally relevant polymorphisms in the human nuclear vitamin D receptor gene. *Mol Cell Endocrinol* **177**, 145-59, 2001
- 12 Jurutka PW, Whitfield GK, Hsieh JC, Thompson PD, Haussler CA, et al.: Molecular nature of the vitamin D receptor and its role in regulation of gene expression. *Rev Endocr Metab Disord* **2**, 203-16, 2001
- 13 Morrison NA, Yeoman R, Kelly PJ, Eisman JA: Contribution of trans-acting factor alleles to normal physiological variability: vitamin D receptor gene polymorphism and circulating osteocalcin. *Proc Natl Acad Sci U S A* **89**, 6665-9, 1992
- 14 Faraco JH, Morrison NA, Baker A, Shine J, Frossard PM: Apal dimorphism at the human vitamin D receptor gene locus. *Nucleic Acids Res* **17**, 2150, 1989
- 15 Falletti E, Bitetto D, Fabris C, Cussigh A, Fontanini E, et al.: Vitamin D receptor gene polymorphisms and hepatocellular carcinoma in alcoholic cirrhosis. *World J Gastroenterol* **16**, 3016-24, 2010
- 16 Morrison NA, Qi JC, Tokita A, Kelly PJ, Crofts L, et al.: Prediction of bone density from vitamin D receptor alleles. *Nature* **367**, 284-7, 1994. doi: 10.1038/367284a0
- 17 Durrin LK, Haile RW, Ingles SA, Coetzee GA: Vitamin D receptor 3'-untranslated region polymorphisms: lack of effect on mRNA stability. *Biochim Biophys Acta* **1453**, 311-20, 1999
- 18 Ingles SA, Wang J, Coetzee GA, Lee ER, Frankl HD, et al.: Vitamin D receptor polymorphisms and risk of colorectal adenomas (United States). *Cancer Causes Control* **12**, 607-14, 2001
- 19 Grundberg E, Lau EM, Pastinen T, Kindmark A, Nilsson O, et al.: Vitamin D receptor 3' haplotypes are unequally expressed in primary human bone cells and associated with increased fracture risk: the MrOS Study in Sweden and Hong Kong. *J Bone Miner Res* **22**, 832-40, 2007. doi: 10.1359/jbmr.070317
- 20 Hung CH, Chiu YC, Hu TH, Chen CH, Lu SN, et al.: Significance of vitamin d receptor gene polymorphisms for risk of hepatocellular carcinoma in chronic hepatitis C. *Transl Oncol* **7**, 503-7, 2014. doi: 10.1016/j.tranon.2014.05.001
- 21 Martin RJ, McKnight AJ, Patterson CC, Sadlier DM, Maxwell AP: A rare haplotype of the vitamin D receptor gene is protective against diabetic nephropathy. *Nephrol Dial Transplant* **25**, 497-503, 2010. doi: 10.1093/ndt/gfp515
- 22 Gezen-Ak D, Dursun E, Bilgic B, Hanagasi H, Ertan T, et al.: Vitamin D receptor gene haplotype is associated with late-onset Alzheimer's disease. *Tohoku J Exp Med* **228**, 189-96, 2012
- 23 Uitterlinden AG, Fang Y, Van Meurs JB, Pols HA, Van Leeuwen JP: Genetics and biology of vitamin D receptor polymorphisms. *Gene* **338**, 143-56, 2004. doi: 10.1016/j.gene.2004.05.014
- 24 Uitterlinden AG, Pols HA, Burger H, Huang Q, Van Daele PL, et al.: A large-scale population-based study of the association of vitamin D receptor gene polymorphisms with bone mineral density. *J Bone Miner Res* **11**, 1241-8, 1996. doi: 10.1002/jbmr.5650110908
- 25 Jenab M, McKay J, Bueno-de-Mesquita HB, van Duijnhoven FJ, Ferrari P, et al.: Vitamin D receptor and calcium sensing receptor polymorphisms and the risk of colorectal cancer in European populations. *Cancer Epidemiol Biomarkers Prev* **18**, 2485-91, 2009. doi: 10.1158/1055-9965.epi-09-0319

- 26 Mahmoudi T, Karimi K, Mohebbi SR, Fatemi SR, Zali MR: Start codon FokI and intron 8 BsmI variants in the vitamin D receptor gene and susceptibility to colorectal cancer. *Mol Biol Rep* **38**, 4765-70, 2011. doi: 10.1007/s11033-010-0613-1
- 27 Wong HL, Seow A, Arakawa K, Lee HP, Yu MC, et al.: Vitamin D receptor start codon polymorphism and colorectal cancer risk: effect modification by dietary calcium and fat in Singapore Chinese. *Carcinogenesis* **24**, 1091-5, 2003. doi: 10.1093/carcin/bgg059
- 28 Slattery ML, Murtaugh M, Caan B, Ma KN, Wolff R, et al.: Associations between BMI, energy intake, energy expenditure, VDR genotype and colon and rectal cancers (United States). *Cancer Causes Control* **15**, 863-72, 2004. doi: 10.1007/s10552-004-1048-6
- 29 Park K, Woo M, Nam J, Kim JC: Start codon polymorphisms in the vitamin D receptor and colorectal cancer risk. *Cancer Lett* **237**, 199-206, 2006. doi: 10.1016/j.canlet.2005.05.048
- 30 Sweeney C, Curtin K, Murtaugh MA, Caan BJ, Potter JD, et al.: Haplotype analysis of common vitamin D receptor variants and colon and rectal cancers. *Cancer Epidemiol Biomarkers Prev* **15**, 744-9, 2006. doi: 10.1158/1055-9965.epi-05-0814
- 31 Kadiyska T, Yakulov T, Kaneva R, Nedin D, Alexandrova A, et al.: Vitamin D and estrogen receptor gene polymorphisms and the risk of colorectal cancer in Bulgaria. *Int J Colorectal Dis* **22**, 395-400, 2007. doi: 10.1007/s00384-006-0163-0
- 32 Gunduz M, Cacina C, Toptas B, Yaylim-Eraltan I, Tekand Y, et al.: Association of vitamin D receptor gene polymorphisms with colon cancer. *Genet Test Mol Biomarkers* **16**, 1058-61, 2012. doi: 10.1089/gtmb.2012.0044
- 33 Li C, Li Y, Gao LB, Wang YY, Zhou B, et al.: Vitamin D receptor gene polymorphisms and the risk of colorectal cancer in a Chinese population. *Dig Dis Sci* **54**, 634-9, 2009. doi: 10.1007/s10620-008-0375-y
- 34 Slattery ML, Sweeney C, Murtaugh M, Ma KN, Caan BJ, et al.: Associations between vitamin D, vitamin D receptor gene and the androgen receptor gene with colon and rectal cancer. *Int J Cancer* **118**, 3140-6, 2006. doi: 10.1002/ijc.21791
- 35 Hughes DJ, Hlavata I, Soucek P, Pardini B, Naccarati A, et al.: Variation in the vitamin D receptor gene is not associated with risk of colorectal cancer in the Czech Republic. *J Gastrointest Cancer* **42**, 149-54, 2011. doi: 10.1007/s12029-010-9168-6
- 36 Parisi E, Rene JM, Cardus A, Valcheva P, Pinol-Felis C, et al.: Vitamin D receptor levels in colorectal cancer. Possible role of BsmI polymorphism. *J Steroid Biochem Mol Biol* **111**, 87-90, 2008. doi: 10.1016/j.jsbmb.2008.05.001
- 37 Xu Y, He B, Pan Y, Deng Q, Sun H, et al.: Systematic review and meta-analysis on vitamin D receptor polymorphisms and cancer risk. *Tumour Biol* **35**, 4153-69, 2014. doi: 10.1007/s13277-013-1544-y
- 38 Ashktorab H, Nguza B, Fatemi M, Nouraie M, Smoot DT, et al.: Case-control study of vitamin D, dickkopf homolog 1 (DKK1) gene methylation, VDR gene polymorphism and the risk of colon adenoma in African Americans. *PLoS One* **6**, e25314, 2011. doi: 10.1371/journal.pone.0025314
- 39 Poynter JN, Jacobs ET, Figueiredo JC, Lee WH, Conti DV, et al.: Genetic variation in the vitamin D receptor (VDR) and the vitamin D-binding protein (GC) and risk for colorectal cancer: results from the Colon Cancer Family Registry. *Cancer Epidemiol Biomarkers Prev* **19**, 525-36, 2010. doi: 10.1158/1055-9965.epi-09-0662
- 40 Kim HS, Newcomb PA, Ulrich CM, Keener CL, Bigler J, et al.: Vitamin D receptor polymorphism and the risk of colorectal adenomas: evidence of interaction with dietary vitamin D and calcium. *Cancer Epidemiol Biomarkers Prev* **10**, 869-74, 2001
- 41 Peters U, Hayes RB, Chatterjee N, Shao W, Schoen RE, et al.: Circulating vitamin D metabolites, polymorphism in vitamin D receptor, and colorectal adenoma risk. *Cancer Epidemiol Biomarkers Prev* **13**, 546-52, 2004
- 42 Peters U, McGlynn KA, Chatterjee N, Gunter E, Garcia-Closas M, et al.: Vitamin D, calcium, and vitamin D receptor polymorphism in colorectal adenomas. *Cancer Epidemiol Biomarkers Prev* **10**, 1267-74, 2001

- 43 Boyapati SM, Bostick RM, McGlynn KA, Fina MF, Roufail WM, et al.: Calcium, vitamin D, and risk for colorectal adenoma: dependency on vitamin D receptor Bsm1 polymorphism and nonsteroidal anti-inflammatory drug use? *Cancer Epidemiol Biomarkers Prev* **12**, 631-7, 2003
- 44 Yamaji T, Iwasaki M, Sasazuki S, Sakamoto H, Yoshida T, et al.: Association between plasma 25-hydroxyvitamin D and colorectal adenoma according to dietary calcium intake and vitamin D receptor polymorphism. *Am J Epidemiol* **175**, 236-44, 2012. doi: 10.1093/aje/kwr295
- 45 Lucock M, Yates Z, Martin C, Choi JH, Boyd L, et al.: Vitamin D, folate, and potential early lifecycle environmental origin of significant adult phenotypes. *Evol Med Public Health* **2014**, 69-91, 2014. doi: 10.1093/emph/eou013
- 46 Lucock M, Yates Z, Boyd L, Naylor C, Choi JH, et al.: Vitamin C-related nutrient-nutrient and nutrient-gene interactions that modify folate status. *Eur J Nutr* **52**, 569-82, 2013. doi: 10.1007/s00394-012-0359-8
- 47 Stephens M, Donnelly P: A comparison of bayesian methods for haplotype reconstruction from population genotype data. *Am J Hum Genet* **73**, 1162-9, 2003. doi: 10.1086/379378
- 48 Stephens M, Smith NJ, Donnelly P: A new statistical method for haplotype reconstruction from population data. *Am J Hum Genet* **68**, 978-89, 2001. doi: 10.1086/319501
- 49 International-Hapmap-Consortium: The International HapMap Project. *Nature* **426**, 789-96, 2003. doi: 10.1038/nature02168
- 50 Abecasis GR, Auton A, Brooks LD, DePristo MA, Durbin RM, et al.: An integrated map of genetic variation from 1,092 human genomes. *Nature* **491**, 56-65, 2012. doi: 10.1038/nature11632
- 51 Morrison NA, George PM, Vaughan T, Tilyard MW, Frampton CM, et al.: Vitamin D receptor genotypes influence the success of calcitriol therapy for recurrent vertebral fracture in osteoporosis. *Pharmacogenet Genomics* **15**, 127-35, 2005
- 52 Kinyamu HK, Gallagher JC, Knezetic JA, DeLuca HF, Prah JM, et al.: Effect of vitamin D receptor genotypes on calcium absorption, duodenal vitamin D receptor concentration, and serum 1,25 dihydroxyvitamin D levels in normal women. *Calcif Tissue Int* **60**, 491-5, 1997
- 53 Choi JH, Yates Z, Veysey M, Heo YR, Lucock M: Contemporary issues surrounding folic Acid fortification initiatives. *Prev Nutr Food Sci* **19**, 247-60, 2014. doi: 10.3746/pnf.2014.19.4.247
- 54 Lucock M, Yates Z: Folic acid fortification: a double-edged sword. *Curr Opin Clin Nutr Metab Care* **12**, 555-64, 2009. doi: 10.1097/MCO.0b013e32833192bc